Background
Diabetes mellitus (DM) accelerates changes in athelosclerosis, which leads to an increase in both clinical incidence of ischemic heart disease and mortality rate [1]. Further, independent of coronary artery disease, type 2 DM has direct adverse effects on the myocardium [2]. To elucidate the mechanisms of these actions, recently, echocardiographical study [3] or some whole heart perfusion experiments using type 2 diabetic model rodents have been performed. Alterations in energy metabolism in the myocardium of type 2 diabetic rodents have been well investigated [4-6], whereas ion homeostasis has been rarely investigated [7] in ischemia-reperfusion study.
Otsuka Long-Evans Tokushima Fatty (OLETF) strain is a spontaneously type 2 DM model rat with diabetic complications [8] that shows late onset hyperglycemia at 18 weeks of age. In the present study, we induced ischemia in perfused OLETF rat hearts in order to examine the changes in pH and proton production, during and after ischemia, as well as the incidence and duration of ventricular arrhythmia after reperfusion. Furthermore, we observed if improvement in diabetic state by troglitazone [9], a thiazolidinedione, acts to the ischemic injury in heart from diabetic OLETF rat.

**Methods**

**Experimental groups**
The present study was undertaken in accordance with the Animals (Scientific Procedures) Act 1986 and conforms with the Guide for the Care and Use of Laboratory Animals published by US National Institutes of Health (NIH Publication No. 85-23, revised 1996).

Male OLETF rats at 6 weeks of age, age-matched non-diabetic Long-Evans Tokushima Otsuka (LETO) rats [8] were obtained from the Otsuka Pharmacology Laboratory (Tokushima, Japan). All of these rats were maintained at the Jikei University animal experiment center and were kept under controlled temperatures (21 ± 2°C) with a 12-hour artificial light and dark cycle. These rats were then divided into 6 subgroups: OLETF at 16 weeks of age (16-O, n = 8), LETO at 16 weeks of age (16-L, n = 8), OLETF at 32 weeks of age (32-O, n = 8), LETO at 32 weeks of age (32-L, n = 8), OLETF at 32 weeks of age treated with troglitazone (32-OT, n = 8), and LETO at 32 weeks of age treated with troglitazone (32-LT, n = 7). Rat standard laboratory chow (type MF, Oriental Yeast Co) was used as the perfusion buffer. Subsequent to the 5 min control perfusion, whole heart ischemia (flow rate 5–10%) was induced by the use of a one-way ball valve which prevented retrograde perfusion during diastole and the hearts were then perfused with electrical pacing (300 beats/min) for 10 min. After reperfusion, the pacing and the ball valve action were stopped for an additional 20 min.

**Body weights**
The body weight (BW) was measured immediately before each heart perfusion experiment.

**Oral glucose tolerance test**
The oral glucose tolerance test (OGTT) was performed in all groups within a week before each perfusion experiment. After 18 hours of fasting, glucose fluid (2 g/kg BW) was administered using a gastric tube. Blood samples were obtained via the tail vein at 3 time points: before the administration of glucose fluid (pre), 60 minutes after loading, and 120 minutes after loading.

**Heart perfusion and ischemia**
Rats in all groups were anesthetized with sodium pentobarbital (50 mg/kg, i.p. injection) and a thoracotomy was performed after 18 hours of fasting. Blood samples were obtained from the inferior vena cava for measurements of glycohemoglobin A1c (HbA1c) levels and serum levels of insulin. The hearts were quickly removed and then immersed in ice-cold Krebs-Henselei bicarbonate solution. The aorta and the pulmonary vein were cannulated, and Langendorff’s retrograde perfusion was initiated at 37°C and a hydrostatic pressure of 80 mmHg. After 10 min, Langendorff perfusion was switched to the physiological perfusion, working heart mode by clamping the aortic inflow line from the Langendorff reservoir, and opening the left atrial inflow and aortic line. The afterload was maintained at 60 mmHg of hydrostatic pressure and the preload was maintained at 7 mmHg throughout the experiment. Modified Krebs-Henselei bicarbonate buffer containing (in mmol) NaCl 118, KCl 4.7, NaHCO3 25, CaCl2 2.5, MgSO4 · H2O 1.2, EDTA 0.5, KH2PO4 1.2 and glucose 11 (37°C, pH 7.4) bubbled with 95% O2 and 5% CO2 was used as the perfusion buffer. Subsequent to the 5 min control perfusion, whole heart ischemia (flow rate 5–10%) was induced by the use of a one-way ball valve which prevented retrograde perfusion during diastole and the hearts were then perfused with electrical pacing (300 beats/min) for 10 min. After reperfusion, the pacing and the ball valve action were stopped for an additional 20 min.

**Hemodynamic measurements and analysis of the coronary effluent**
The aortic flow was measured with an electromagnetic flowmeter (Nihon-Koden MFV 2100, Tokyo). An 18-gauge catheter was inserted via the left atrium into the left ventricle to measure the left ventricular (LV) pressure, the peak positive first derivative of LV pressure (LV +dP/dt) and peak negative first derivative of LV pressure (LV -dP/dt) with a polygraph system (Fukuda Denshi Co., Tokyo, MIC 8600). An electrocardiogram (ECG) recorded from an epicardium was monitored using carbon lead attached to the surface of the heart. The coronary flow was collected by a heart chamber at 5 min intervals throughout the period of perfusion. Cardiac output was estimated as the sum of the aortic and coronary flow.

The coronary effluent was collected from the pulmonary artery and used for determination of venous PO2, PCO2, HCO3- and pH levels with a blood gas analyzer (Corning 175, USA).

**Arrhythmia study**
ECG was continuously recorded on a recorder throughout the experiment. The ECG data were retrospectively analyzed, in a blind manner, for the incidence, time to onset, and duration of ventricular fibrillation (VF) during reperfusion. All analyses were carried out in accordance with the Lambeth Conventions [10]. VF was defined as a signal in which individual QRS deflections could no longer be
distinguished from one another and the heart rate could not be determined.

Biochemical analysis

The blood glucose level was measured in the whole blood using glucose oxidase method. HbA1c was measured by the latex method. The plasma insulin level was measured using the enzyme-immunoassay insulin kits (Morinaga, Tokyo Japan).

The proton production in coronary effluent was calculated from the $PCO_2$ and $HCO_3^-$ values as follows: $[H^+]$ (n mol/l) $= 24 \times PCO_2$(mmHg)/[$HCO_3^-$] (m mol/l)

Statistical analysis

Values are expressed as the mean ± standard error. The data were analyzed using either Student's t-test for unpaired data or analysis of variance followed by appropriate post-hoc test to locate differences between groups. Binominal distributed variables, such as the incidence of VF, were compared using the $\chi^2$ test for a 2 × n table, followed by a sequence of 2 × 2 $\chi^2$ tests with Yates's correction. A value of p < 0.05 was considered to be statistically significant.

Results

Characteristics of OLETF and LETO rats

Body weight was significantly higher in 16-O compared with that in 16-L, while 32-O seemed to be obese, with body weight significantly higher as compared with 32-L and 16-O. No significant differences were seen in body weights among the groups based on troglitazone treatment. The level of HbA1c showed no difference between 16-L and 16-O, however, that in 32-O was significantly higher than that in 32-L. HbA1c in 32-OT was significantly lower as compared with 32-O, while there was no significant difference between 32-LT and 32-L. The plasma glucose level in 16-O was higher than that in 16-L before OGTT, however, there were no differences between those 2 groups in regard to OGTT results. The level of plasma glucose in 32-O showed a significantly higher level than that in 32-L before and during OGTT, while a significant improvement in plasma glucose level in 32-OT was observed following treatment with troglitazone before and in OGTT results. In contrast, troglitazone did not have an affect on plasma glucose level in 32-LT. The serum insulin level showed no significant difference between 16-O and 16-L, however, that in 32-O tended to be higher than that in 32-L (p= 0.08). Hyperinsulinemia seen in 32-O improved slightly, as shown in 32-OT (p = 0.10), by treatment with troglitazone. These results are shown in Table 1.

Basal cardiac function

In the perfused working hearts, there were no differences in the heart rate, coronary flow, cardiac output, LV pressure, and LV ± dP/dt between 16-O and 16-L. The effects of aging in these parameters were not seen between 32-O and 16-O, and between 32-L and 16-L. In addition, there were no significant differences in these parameters between 32-OT and 32-O, or between 32-LT and 32-L. These results are shown in Table 2.

Reperfusion induced arrhythmia

Reperfusion induced arrhythmia VF occurred during the early phase of reperfusion after ischemia. There were no significant differences in the incidence of VF between OLETF and age-matched LETO rats. The incidence of VF (Fig. 1A) was 50% in both 16-O and 16-L, while that in 32-O was 100%, though it decreased (32-OT: 50%, p = 0.10) by treatment with troglitazone. There was no significant difference in the duration of VF (Fig. 1B) between 16-O and 16-L. That in 32-O was significantly longer than in 32-L. The duration of VF in 32-O was significantly shortened by treatment with troglitazone, and that in 32-L was also significantly shortened as compared with 32-LT.

Changes in pH of coronary effluent

In all groups, pH of the coronary effluent decreased during ischemia and then recovered after ischemia. There were no significant differences in pH values of coronary effluent samples throughout the ischemia-reperfusion

### Table 1: Characteristics of the LETO and OLETF rats in each groups

<table>
<thead>
<tr>
<th></th>
<th>16-L (n = 8)</th>
<th>16-O (n = 8)</th>
<th>32-L (n = 8)</th>
<th>32-O (n = 8)</th>
<th>32-LT (n = 7)</th>
<th>32-OT (n = 8)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body Weight (g)</td>
<td>383.8 ± 8.9</td>
<td>468.8 ± 10.6***</td>
<td>460.8 ± 7.6</td>
<td>563.2 ± 15.2††</td>
<td>465.0 ± 14.9</td>
<td>521.4 ± 21.5</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>3.04 ± 0.07</td>
<td>3.18 ± 0.06</td>
<td>3.17 ± 0.07</td>
<td>4.45 ± 0.26†</td>
<td>3.09 ± 0.13</td>
<td>3.52 ± 0.07#</td>
</tr>
<tr>
<td>Serum Insulin (ng/ml)</td>
<td>1.5 ± 2.3</td>
<td>2.4 ± 0.8</td>
<td>1.7 ± 0.9</td>
<td>3.3 ± 1.1</td>
<td>1.6 ± 0.6</td>
<td>1.8 ± 0.2</td>
</tr>
<tr>
<td>Plasma glucose in OGTT (mg/dl)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>pre</td>
<td>86.6 ± 2.2</td>
<td>100.9 ± 4.1*</td>
<td>106.3 ± 3.7</td>
<td>122.3 ± 5.1†</td>
<td>95.8 ± 2.0</td>
<td>98.8 ± 3.9#</td>
</tr>
<tr>
<td>60 min</td>
<td>131.1 ± 4.1</td>
<td>157.1 ± 14.4</td>
<td>142.5 ± 4.1</td>
<td>310.7 ± 5.0††</td>
<td>134.9 ± 1.7</td>
<td>225.6 ± 6.9#</td>
</tr>
<tr>
<td>120 min</td>
<td>102.6 ± 4.1</td>
<td>100.9 ± 1.9</td>
<td>101.9 ± 1.9</td>
<td>161.5 ± 4.8††</td>
<td>95.1 ± 3.1</td>
<td>98.9 ± 4.3#</td>
</tr>
</tbody>
</table>

Values are the means ± SEM. HbA1c, glycohemoglobin A1c; OGTT, oral glucose tolerance test.

* p < 0.05 vs 16-L, **p < 0.01 vs 16-L, †p < 0.05 vs 32-L, ††p < 0.01 vs 32-L, †p < 0.05 vs 32-O, #p < 0.05 vs 32-O, ##p < 0.01 vs 32-O
protocol between 16-O and 16-L (Fig. 2A), however, pH in 32-O was significantly lower than in 32-L at 9 minutes during ischemia (Fig. 2B). The pH value in 32-LT did not show a significant difference compared with that in 32-L throughout the ischemia-reperfusion protocol (Fig. 3A), though the decrease in pH was significantly suppressed in 32-OT as compared with 32-O at 5 and 9 minutes during ischemia (Fig. 5B).

Changes in proton production in coronary effluent
Proton efflux increased during ischemia and recovered after reperfusion in all groups. No significant difference was observed between the proton efflux levels in 16-O and 16-L (Fig. 4A), whereas that in 32-O was significantly higher than that in 32-L at 5 and 9 minutes during ischemia (Fig. 4B). No significant difference was observed between 32-LT and in 32-L throughout the perfusion protocol (Fig. 5A), while the increase in proton efflux was significantly suppressed in 32-OT as compared with 32-O at 5 and 9 minutes during ischemia (Fig. 5B).

Table 2: Myocardial function before ischemia in each group.

<table>
<thead>
<tr>
<th></th>
<th>16-L (n = 8)</th>
<th>16-O (n = 8)</th>
<th>32-L (n = 8)</th>
<th>32-O (n = 8)</th>
<th>32-LT (n = 7)</th>
<th>32-OT (n = 6)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HR (beats/min)</td>
<td>212.3 ± 13.2</td>
<td>202.5 ± 7.7</td>
<td>219.1 ± 7.7</td>
<td>202.8 ± 10.7</td>
<td>218.2 ± 5.6</td>
<td>201.0 ± 9.3</td>
</tr>
<tr>
<td>CF (ml/min)</td>
<td>16.0 ± 1.0</td>
<td>17.1 ± 1.0</td>
<td>19.5 ± 0.5</td>
<td>21.1 ± 0.8</td>
<td>21.2 ± 0.7</td>
<td>21.7 ± 0.4</td>
</tr>
<tr>
<td>CO (ml/min)</td>
<td>45.1 ± 3.5</td>
<td>52.9 ± 2.2</td>
<td>51.1 ± 3.0</td>
<td>50.4 ± 4.0</td>
<td>58.1 ± 2.7</td>
<td>51.0 ± 2.8</td>
</tr>
<tr>
<td>LVP (mmHg)</td>
<td>133.0 ± 3.0</td>
<td>142.8 ± 4.6</td>
<td>135.5 ± 4.9</td>
<td>134.1 ± 6.3</td>
<td>140.1 ± 1.9</td>
<td>130.7 ± 2.8</td>
</tr>
<tr>
<td>LV +dP/dt (mmHg/sec × 10^3)</td>
<td>2.6 ± 0.3</td>
<td>2.9 ± 0.2</td>
<td>4.0 ± 0.2</td>
<td>3.6 ± 0.2</td>
<td>4.1 ± 0.1</td>
<td>3.4 ± 0.1</td>
</tr>
<tr>
<td>LV -dP/dt (mmHg/sec × 10^3)</td>
<td>2.5 ± 0.2</td>
<td>2.5 ± 0.3</td>
<td>3.2 ± 0.3</td>
<td>2.8 ± 0.3</td>
<td>3.3 ± 0.3</td>
<td>3.0 ± 0.2</td>
</tr>
</tbody>
</table>

Values are mean SEM. HR, Heart rate; CF, Coronary flow; CO, Cardiac output; LVP, Left ventricular pressure; LV +dP/dt, Peak positive first derivative of left ventricular pressure; LV -dP/dt, Peak negative first derivative of left ventricular pressure.

Figure 1
Incidence (A) and duration (B) of ventricular fibrillation after reperfusion. * p < 0.05 vs. OLETF rats at 32 weeks of age. † p < 0.05 vs. LETO rats at 32 weeks of age.

Figure 2
Changes in pH of coronary effluent in LETO and OLETF rats at 16 weeks of age (A) and 32 weeks of age (B). Closed circles show LETO rats, open circles show OLETF rats. * p < 0.05 vs. LETO rats.
Discussion

In the present study, a prolonged duration of reperfusion arrhythmia was observed in hearts from OLETF rats at 32 weeks of age, in which obesity and hyperglycemia were confirmed by OGTT results. Further, pH levels were lower and proton production was significantly higher in the coronary effluent of those rats as compared with age matched LETO rats. In addition, the duration of reperfusion arrhythmia was significantly shortened, while the decrease in pH and increase in proton production were significantly suppressed in coronary effluent samples following treatment with troglitazone in the hearts of those OLETF rats.

In the present study, the plasma glucose level at 60 min during OGTT was greater than 300 mg/dl in OLETF rats at 32 weeks of age, and the HbA1c level was also significantly higher as compared to the age matched LETO rats. According to our results as well as those of other studies [8,11], OLETF rats at 32 weeks of age in the present study were at the impaired glucose tolerance (IGT) stage. Demonstrating IGT and obesity, these characteristics resembled human type 2 DM at an early stage. The plasma glucose levels after fasting and in the OGTT results, as well as HbA1c and serum insulin levels, were improved in OLETF rats at 32 weeks of age by treatment with troglitazone as in other reports [12].

There have been other studies on cardiac function in diabetic OLETF rats. Mizushige et al. [3] reported the occurrence of a left ventricular diastolic dysfunction in the prediabetic stage of OLETF rat using Doppler echocardiography. Abe et al. [13] also reported occurrence of left ventricular diastolic dysfunction in diabetic OLETF rats at 62–66 weeks of age in experiments with isolated hearts, though that dysfunction was not observed at 40–46 weeks
of age. Impaired cardiac contraction during working heart perfusion in OLETF rats at 12 months of age [4] and an impairment in vivo cardiac contraction in OLETF rats at 36 [14] and 62 [15] weeks of age have also been reported. In the present study, the LV functional parameters before ischemia did not differ between the LETO and OLETF rats at 32 weeks of age. We considered that the discrepancies between these reports may have been caused by the different experimental methods and stage of diabetes in the animals. Meanwhile, one of the fatty obese rodents, ICR: LAC-p rats have been reported to have normal basal myocardial function, while those animals also exhibited an increased sensitivity to ischemic myocardial injury that developed with advancing age [5]. The present results for OLETF rat hearts are similar to those findings.

The pH values in coronary effluent did not drop dramatically after inducing ischemia, in contrast to myocardial intracellular pH levels measured in other studies of no-flow ischemia [6,16]. In the present study, low-flow ischemia was induced, which might be the reason for this discrepancy. However, pH values decreased significantly after 10 min of ischemia as compared with those before ischemia in all groups (Figure 2 and 3). Additionally, those decreases in pH values after 10 min of ischemia (∆pH = 0.3–0.4) were similar to the results of other reports with low-flow ischemia [16]. Thus, the most important finding regarding cardiac metabolism in the present study was exacerbated acidosis during ischemia in the hearts of OLETF rats after the incidence of IGT. As sources of acidosis during ischemia in hearts, anaerobic glycolysis, CO₂ production from respiration in mitochondria, impairment in the oxygenation of NADH₂, and the effects of proton production cycle are well-known. We previously reported higher levels of lactate production during ischemia in the hearts of OLETF rats at 32 weeks of age [17]. Thus, the findings in the present study may mainly correlate with an acceleration of anaerobic glycolysis during ischemia in the myocardium of OLETF rats at that age. Anaerobic glycolysis is an important source of ATP production, however, it causes increases in the accumulation of acidic metabolites and decreases in intracellular pH, and is also correlates with the severity of myocardial damage. In addition, local acidosis leads to alterations in electrical currents and electrophysiological changes in cell membranes [18]. While, severe acidosis and elevated proton production increase intracellular Na⁺ via Na⁺-H⁺ exchanger during ischemia in the myocardium. Increased intracellular Na⁺ causes intracellular Ca²⁺ overload after reperfusion, and reperfusion arrhythmia occurs [19]. Regarding these previous reports, exacerbation of acidosis and increased proton production can be correlated with prolonged duration of reperfusion induced VF in the hearts of OLETF rats at 32 weeks of age. Additionally, IGT improved, and exacerbation of acidosis and an increase in proton production in the coronary effluent were suppressed during ischemia at that age by treatment with troglitazone. Moreover, the duration of reperfusion induced VF was shortened simultaneously. These results confirmed the correlation between impaired glucose metabolism and increased sensitivity to ischemia in the myocardium of OLETF rats after the incidence of IGT. In ischemia-reperfusion experiments using spontaneous type 2 diabetic rodent hearts, impaired recovery of cardiac function after reperfusion has been reported [4]. In a heart perfusion study conducted by Maddaford et al. [5], ATP and glycogen contents in the insulin-resistant rat myocardium decreased significantly after ischemia-reperfusion, indicating acceleration of the consumption and drying of stored energy. Sidell et al [6] reported a greater loss of ATP and lactate production in the Zucker Fatty rat hearts during ischemia, thus correlating to a lower recovery in the contractile function during reperfusion. In the only known report of reperfusion arrhythmia in spontaneous type 2 diabetic animal hearts, which utilized db/db mice, an increase in the incidence and prolonged duration of reperfusion arrhythmia were observed [7]. In that study, a greater increase in intracellular Na⁺ was indicated as the main cause of those conditions. As for ion homeostasis, altered intracellular Ca²⁺ handling [20] and Kᵦᵦ current [21] in spontaneous diabetic mouse myocytes have been reported. These alterations also have possibility to increase the incidence or prolong the duration of reperfusion arrhythmia. Thus, some metabolic mechanisms or ion homeostasis apart from intracellular acidosis and proton efflux may correlate with increased sensitivity to ischemia in hearts of OLETF rats at 32 weeks of age.

We also observed a shortening of the duration of reperfusion arrhythmia VF following treatment with troglitazone in the hearts of non-diabetic LETO rats at 32 weeks of age. There was not a significant difference in pH level and proton efflux between the treated and non-treated groups. Then, some mechanisms apart from improvement of glucose metabolism might shorten the duration of the reperfusion induced VF in the hearts of OLETF rats at 32 weeks of age treated with troglitazone. In fact, an improvement in the recovery of cardiac function after ischemia has been reported in non-diabetic pig hearts treated with troglitazone for a long time [22]. However, after treatment with troglitazone, the incidence of VF did not improve in the hearts of LETO rats at 32 weeks of age, whereas that in the hearts of age matched OLETF rats tended to be improved (p = 0.10). These results demonstrate that suppression of alteration in glucose metabolism according to the incidence of IGT in myocardium should be the main mechanism improving the exacerbation of reperfusion injury by treatment with troglitazone in OLETF rat. Recent clinical trials have revealed that thiazolidinediones provide a better prognosis in diabetic
patients with coronary heart disease [23]. However, the effects of thiazolidinediones, including troglitazone, on the heart have not been well clarified and additional investigations are required.

Conclusion
Sensitivity to ischemia increased after the incidence of impaired glucose tolerance in the hearts of OLETF rats, which improved by the treatment with troglitazone. We concluded that exacerbation of acidosis through altered glucose metabolism during ischemia is one of the causes of this phenomenon in diabetic OLETF rats.

Abbreviations
DM, diabetes mellitus; OGTT, oral glucose tolerance test; VF, ventricular fibrillation.

Acknowledgements
The authors thank Sankyo Co, Ltd. for providing the troglitazone.

References

Publish with BioMed Central and every scientist can read your work free of charge

BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime.
Sir Paul Nurse, Cancer Research UK

Your research papers will be:
- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

Page 7 of 7
(page number not for citation purposes)