PGF2alpha induced differential expression of genes involved in turnover of extracellular matrix in rat decidual cells
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Abstract

In the rat, the decidual tissue is an important component for maternal recognition of pregnancy. Decidualization can be induced by either the implantation of the blastocyst or by artificial stimuli. The process of decidua formation or decidualization, is characterized by growth and differentiation of endometrial stromal cells. Prostaglandin F2alpha (PGF2α) has been shown to be involved in inhibition of implantation, alteration of embryo development, induction of luteal regression, and the mediation of pregnancy loss induced by microorganism infections. In order to establish a direct role for PGF2α in decidual function, we have evaluated its effects on the expression of an extensive array of genes using primary decidual cell culture. Upon treatment with PGF2α sixty genes were significantly down-regulated whereas only six genes were up-regulated (from a total of 1176 genes studied). Interestingly, the majority of the genes inhibited by PGF2α are either directly or indirectly involved in the turnover of the extracellular matrix (ECM). Genes such as gelatinase A (MMP2), cathepsin L, tissue inhibitor metalloproteinases 2 (TIMP2) and 3 (TIMP3), plasminogen activator inhibitor1 (PAI1), tissue type plasminogen activator (tPA), urokinase plasminogen activator (tPA), endothelin 1, calponin, carboxypeptidase D and calponin acidic were down regulated. The opposite effect was observed for prostromelysin 53 kDa (proMMP3), plasma proteinase I alpha and alpha 1 antiproteinase, all of which were significantly up-regulated by PGF2α. The results strongly suggest that the abortificient role of elevated levels of PGF2α after implantation is due, in large part, to inhibition of genes involved in the normal turnover of the extracellular matrix necessary for decidual formation.

Background

The establishment of successful pregnancy requires a profound reorganization of uterine tissues. Rapid growth and differentiation of the endometrial stroma is the earliest and most striking event in pregnancy. Differentiation of stromal cells leads to the formation of unique cells, termed decidual cells, which differ greatly from the original stromal cells [1]. The decidua is an important component in the maternal recognition of pregnancy and can be induced by either the implantation of the blastocyst or by artificial stimuli. An interesting feature is that – the growth and differentiation of the endometrial cells – occurs differently in different regions of the uterus [1]. Mesometrial decidual cells are formed after the antimesometrial decidua and their death takes place after antimesometrial cell degeneration. Regression of the decidual cells appears
to be controlled by an intrinsic cell death program of apoptosis, which takes place after day 10 of pregnancy in the antimesometrial region first, followed by the mesometrial region [2].

Prostaglandin F$_{2\alpha}$ can induce different biological actions at the beginning and at the end of pregnancy. PGF$_{2\alpha}$, Prostaglandin E$_2$ (PGE$_2$), and 6-keto-PGF$_{1\alpha}$ are produced by the pregnant uterus [3]. An increase of uterine PGE$_2$ and PGF$_{2\alpha}$ is observed on day 5 of pregnancy, allowing the decidualization process to take place. When embryo access to the uterus is impaired, production of prostaglandins (PGE$_2$ and PGF$_{2\alpha}$) is suppressed [4]. During postimplantation, PGE2 returns to the original preimplantation levels, but PGF$_{2\alpha}$ decreases [4]. Whereas PGF2α contributes to the process of decidualization, implantation and recognition of pregnancy. An increase in PGF$_{2\alpha}$ over certain values can terminate pregnancy [6]. A high level of PGF$_{2\alpha}$ is known to induce inhibition of implantation, alteration of embryo development, and induction of luteal regression [5]. During infection, an inflammatory response mediated by cytokines can be generated [7]. This release of PGF$_{2\alpha}$ can induce premature uterine contraction and premature labor [8]. Depending on which stage of pregnancy an increase of PGF$_{2\alpha}$ secretion occurs, it can alter implantation, induce abortion or even embriolethality [7,8]. Because an increase in PGF$_{2\alpha}$ levels after implantation can be detrimental for the progression of pregnancy, the aim of this investigation was to determine whether PGF$_{2\alpha}$ affects directly decidual cells and leads to disturbance in the expression of genes crucial for decidual survival.

Methods

Animal model

Pseudopregnancy was induced by mating Holtzman Sprague Dawley female rats with vasectomized male rats. The day a vaginal plug was found was designated day 1 of pseudopregnancy. Decidualization of the uterine endometrium was induced by scratching the antimesometrial surface with a hooked needle on day 5 of pseudopregnancy under ether anaesthesia. Animals were housed in a controlled environment (22–24°C) and kept under controlled conditions (lights on: 0500–1900 hrs) with free access to standard rat chow and water. The University of Illinois at Chicago animal care and use committee approved the animal care and handling.

Primary decidual cell culture and hormone treatment

For each experiment 15 pseudopregnant rats were used. Decidual cells were isolated as previously described [9]. Cells were pooled and seeded into six-well plates (1.4 × 10$^6$ cells/ well). They were allowed to attach 3–4 hrs before washing in PBS to remove erythrocytes. Cells were incubated in RPMI 1640 without phenol red (Mediatech, Whashington DC), containing 1% CD-FBS (HyClone Laboratories Inc, Logan, UT), and treated with or without high levels (5 μM [10,11]) of PGF$_{2\alpha}$ (Sigma, St. Louis, MO) for 12 hr. After incubation, the cells were harvested in cold PBS, quick frozen in liquid nitrogen and kept at -80°C until RNA isolation.

Gene identification by cDNA array

Total RNA was isolated from primary decidual cells (PDC) by using a RNAII isolation kit (Clontech, Palo Alto, CA), following the manufacture's instructions. RNA isolated from wells treated with either PGF$_{2\alpha}$ or vehicle were pooled independently and subjected to cDNA array. cDNA probes were generated from 4 μg of total RNA in a reverse transcriptase reaction using a mix of dATP, dTTP and dGTP (Takara Biomedicals, Shiga, Japan) plus [α$^{32}$P]-dCTP (Amersham, MO), and a mixture of primers specific to each gene present in the array. All probes used had 5 to 10 × 10$^6$ cpm and the difference between control and experimental probes in each assay was less than 10%. cDNA were hybridized to an Atlas Rat 1.2 Array (#7854-1) nylon membrane (Clontech, CA). Hybridization and post-hybridization washes were performed according to the manufacturer's protocol. The signal was scanned with a phosphorImager ( Molecular Dynamics, CA) after 48 h of exposure. Control and experimental RNA were always processed in parallel.

Data analysis

Spot intensities from scanned membranes were analyzed using the AtlasImage 1.5 software (Clontech, CA). Grids were orientated manually and adjusted to ensure optimal spot recognition using AtlasImage's fine tuning tools, discarding spots with dust or locally high background. The software makes the analysis for background correction and normalization versus housekeeping genes. It also calculate the ratio and indicates which genes has a ratio > 2 (up regulated), or ratio < 0.5 (down regulated) or 0.5 < ratio < 2 (equal expression). Gene expression data were normalized using the Sum method included in the Atlas Image software. Data points where the expression was not greater than two standard deviations were discarded. For the final analysis, data points were the averages from triplicates and any non-reproducible data were discarded. The relative RNA expression with differences between control and treatment being higher or equal to 2 and lower than 0.5 were considered significantly different [12].

Results

Effect of PGF$_{2\alpha}$ on gene expression in primary decidual cells

From the total genes available in the rat 1.2 membrane arrays, only 20% of the genes were detected in rat primary decidual cells. Sixty genes were significantly down regulated, whereas only six genes were up regulated by PGF$_{2\alpha}$.
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**Effect of PGF$_{2\alpha}$ on the expression of genes related to the extracellular matrix (ECM)**

The majority of genes whose expression was affected by PGF$_{2\alpha}$ in primary decidual cells are genes involved in the regulation of the ECM. Genes such as gelatinase A (MMP2), cathepsin L, tissue inhibitor metalloproteinases 2 (TIMP2) and 3 (TIMP3), plasminogen activator inhibitor 1 (PAI1), tissue type plasminogen activator (tPA), urokinase plasminogen activator (uPA), endothe- lin 1, calponin, carboxypeptidase D and calponin acidic were down regulated. The opposite effect was observed for prostromelysin 53 kDa (proMMP3), plasma proteinase inhibitor 1 alpha and alpha 1 antiproteinase - all of which were significantly up regulated by PGF$_{2\alpha}$ (Table 1).
Figure 2
cDNA expression array of rat primary decidual cells treated with PGF$_{2\alpha}$ or vehicle.
PGF2α regulation of genes related to the proteasome component system

Messenger RNA encoding proteasome Iota, proteasome component C2, proteasome subunit RC7-I, 26-S proteasome regulator and proteasome component C3 were down regulated by PGF2α. Only proteasome component C9 was up regulated after PGF2α treatment (Table 2).

PGF2α, regulation of genes involved in trafficking and signal transduction

The mRNAs encoding for 14-3-3ε and z/δ proteins, and Annexin IV were down regulated in PDC after PGF2α treatment (Table 3). PGF2α down regulated the expression of serum glucocorticoid kinase (SGK), casein kinase I, extracellular signal-regulated kinase (ERK-1), male germ cell-associated kinase (MAK), and Wee tyrosine kinase. PGF2α also significantly inhibited Crk-associated kinase (CAS), calcium calmodulin kinase II (CAMKII) and IV (CAMKIV), phospholipase C1, and protein phosphatases such as phosphatase 2A and protein tyrosine phosphatase 1B (Table 3).

| Table 1: Effect of PGF2α on the expression of genes related to the extracellular matrix (ECM). |
|---------------------------------|----------------|----------------|----------|----------------|----------------|----------|----------|
| **Gene Bank number** | **Swiss protein number** | **Name** | **Function** | **Ratio ± SEM** | **% decrease** | **Fold increase** |
| U65656 | P97581 | Gelatinase A | Metalloproteinase | 0.04 ± 0.02 | 87.53 |
| Y00697 | P07154 | Cathepsin L | Cysteine protease | 0.09 ± 0.05 | 91.09 |
| L31884 | P30121 | Tissue inhibitor metalloproteinase 2 (TIMP2) | Protease inhibitor | 0.05 ± 0.01 | 95.54 |
| U27201 | P48032 | Tissue inhibitor metalloproteinase 3 (TIMP3) | Protease inhibitor | 0.04 ± 0.01 | 91.95 |
| X63434 | P29598 | Urokinase type Plasminogen activator (uPA) | Serine protease | 0.14 ± 0.09 | 94.86 |
| M24067 | P20961 | Plasminogen activator inhibitor (PAI) | Protease inhibitor | 0.09 ± 0.05 | 91.68 |
| M2367 | P19367 | Tissue type plasminogen activator (tPA) | Serine protease | 0.17 ± 0.02 | 89.90 |
| X71071 | Q08290 | Calponin | Cytoskeleton | 0.08 ± 0.004 | 92.50 |
| U06755 | P37397 | Calponin, acidic | Cytoskeleton | 0.24 ± 0.13 | 76.76 |
| U62897 | O35850 | Carboxypeptidase D | Carboxypeptidase | 0.24 ± 0.14 | 76.48 |
| M64711 | P22388 | Endothelin I | Hormone | 0.34 ± 0.2 | 77.48 |
| X02601 | P03957 | Polypeptide, 53 kDa, growth factor induced | Metalloproteinase | 2.54 ± 1.04 | 3 |
| J03552 | P14046 | Plasma protease inhibitor α1-inhibitor III | Protease inhibitor | 3.64 ± 0.64 | 4 |
| M32247 | P17475 | α1 antiprotease | Protease inhibitor | 2.45 ± 0.44 | 3 |

PGF2α regulation of genes related to the proteasome component system

Table 2: PGF2α regulation of genes related to the proteasome component system.

<table>
<thead>
<tr>
<th><strong>Gene Bank number</strong></th>
<th><strong>Swiss Protein number</strong></th>
<th><strong>Name</strong></th>
<th><strong>Function</strong></th>
<th><strong>Ratio ± SEM</strong></th>
<th><strong>% decrease</strong></th>
<th><strong>Fold increase</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>D10755</td>
<td>P34062</td>
<td>Proteasome Iota</td>
<td>Proteasomal protein</td>
<td>0.07 ± 0.04</td>
<td>93.64</td>
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<tr>
<td>M29859</td>
<td>P18420</td>
<td>Proteasome component C2</td>
<td>Proteasomal protein</td>
<td>0.31 ± 0.15</td>
<td>78.28</td>
<td></td>
</tr>
<tr>
<td>U77918</td>
<td>Q63569</td>
<td>26-proteasome component C3</td>
<td>Proteasomal protein</td>
<td>0.39 ± 0.08</td>
<td>61.94</td>
<td></td>
</tr>
<tr>
<td>J03897</td>
<td>P17220</td>
<td>Proteasome component C3</td>
<td>Proteasomal protein</td>
<td>0.31 ± 0.02</td>
<td>59.02</td>
<td></td>
</tr>
<tr>
<td>D21799</td>
<td>P40307</td>
<td>Proteasome subunit RC7-I</td>
<td>Proteasomal protein</td>
<td>0.18 ± 0.01</td>
<td>82.62</td>
<td></td>
</tr>
<tr>
<td>X55986</td>
<td>P21670</td>
<td>Proteasome subunit C9</td>
<td>Proteasomal protein</td>
<td>2.32 ± 0.72</td>
<td>3</td>
<td></td>
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</tbody>
</table>

Ratio = net intensity of PGF2α normalized/net intensity of control normalized, and represent the mean ± SEM of each ratio from separate experiments. Ratio > 2 = up regulation; ratio < 0.5 = down regulation; 0.5 < Ratio < 2 = equal expression.
cytokine and chemokine related genes found significantly down regulated in response to PGF2α. Conversely, PGF2α stimulated the expression of basic fibroblast growth receptor factor 1 (bFGFRF1) (Table 4).

**Discussion**

Levels of PGF2α in the uterus need to be under tight control to avoid interference with the establishment and progression of pregnancy. Pathophysiological elevations in PGF2α lead to excessive uterine contractions. Therefore, PGF2α production must be avoided to maintain a quiescent uterus [13]. In this paper we present initial data demonstrating that elevated PGF2α can target the decidua, an endocrine tissue whose integrity is fundamental for the success of implantation and for the progression of pregnancy. Our results demonstrate that elevated PGF2α down regulate the expression of decidual genes related to the proteasome protein component system, those involved in trafficking and signal transduction and genes associated with G-proteins, growth factors, chemokines and cytokines. However, interestingly, the main effect of PGF2α is related to the turnover and degradation of the ECM which provides mechanical strength to the tissue. Thus, in addition to inducing uterine contractions, PGF2α can have a noxious impact on the success of implantation and progression of pregnancy, at least in part by deregulating the turnover of the ECM in the decidua.

The largest group of genes down regulated by PGF2α is directly or indirectly related with the turnover and degradation of the ECM. Genes such as gelatinase A (MMP-2), TIMP2 and 3 [14,15], cathepsin L [16], carboxypeptidase D are included in different groups of proteases, and can directly affect the degradation of the ECM. Carboxypeptidase D (CPD) or metallocarboxypeptidase D is a 180-kDa protein that contains almost three carboxypeptidase-like domains, a transmembrane domain, and a cytosolic tail. This gene participates in the processing of proteins transiting the secretory pathway [17]. TGF-β is a key factor that favours accumulation of collagen, laminin and fibronectin in the ECM [17]. TGF-β can stimulate PAI-1, inhibiting the protease degradation of the ECM. Other factors included in the plasminogen/plasmin and fibrinolytic system, such as IPA, uPA and PAI-1, can participate in the tissue remodelling of the ECM directly through the binding to specific receptors, and through the transformation of the plasminogen precursor bound to the cell surface to plasmin, which is an active serine protease. Plasmin is able to degrade most of the components of the ECM either directly or indirectly by the activation of MMPs. All of the plasminogen/plasmin factors mentioned previously participate in the process of decidualization [19-21].

Endothelin-1 is a peptide characterized as a potent endothelial cell-derived vasoconstrictor. It is synthesized as an inactive precursor (preproendothelin) and processed to a mature active form (endothelin) by zinc metalloproteinases. The active form, endothelin-1, promotes synthesis of collagen types I and II by fibroblasts, affects the ECM remodelling and promotes the proliferation of mesangial cells [21]. Endothelin-1 is also associated with neovascularization and regulation of blood flow [22]. This vasoactive factor is also involved in the genesis of endothelial cells behaviour [22,23].

Annexin IV (ANX4, also called Lipocortin) was differentially down regulated by PGF2α. This protein belongs to a
family of intracellular proteins that binds membrane phospholipids in a calcium-dependent manner. Thus, it can also inhibit phospholipase A2 (PLA2) [24]. ANX4 can also directly bind glycosaminoglycan (GAG) and can be localized not only to the cytoplasm but also the cell surface or the extracellular compartment [25]. It has been proposed that ANX4 could affect the mobilization of different substrates involved in the regulation of the ECM. Moreover, Annexin IV and other annexins can act as ligands for proteoglycans localized on the cell surface, in the ECM, or in secretory granules [24,25].

Calponin (CaP) has three genetic isoforms, h1, h2 and acidic calponin, and can be identified by the individual C-terminal tail sequences. The c-terminal sequences regulate actin association and the cytoskeleton [26,27]. It is known that Cap or basic Cap inhibit the actomyosin ATPase in a calmodulin dependent manner [27]. Basic Cap, by affecting microtubules assembly, can modify the cytoskeleton of the cells, and indirectly, the associated ECM [27,28]. On the other hand, acidic Cap belongs to the family of actin-associated proteins. It can interact with F-actin but not with microtubules, desmin filaments, and tropomyosin as basic Cap does. These properties suggest that acidic Cap is functionally distinct from basic Cap and could affect the ECM in a different manner [29].

Pro-stromelysin, α1 antiproteinase, plasma proteinase-1α inhibitor, basic fibroblast growth receptor factor 1, phosphorylase B and proteasome component C9 were the only genes up regulated by PGF2α in PDC. Pro-stromelysin is a 53 kDa peptide and a pro-MMP3 precursor and is directly related with the ECM regulation [30]. Plasma proteinase 1α inhibitor belongs to a major group of proteins that includes α2-macroglobulin [31], an important protein secreted by the decidual cells which has a critical role in the control of implantation [31]. Growth factor receptors, such as bFGFR, are part of a multigene family of structurally related factors (FGFs 1 to 9), some of which bind heparin sulphate proteoglycans that are components of the ECM [32]. bFGFRs as well as bFGF are also temporally and spatially present in the pregnant rat uterus [33-35]. Another member of the TGFβ superfamily expressed in the uterus during early implantation and down-regulated by PGF2α was BMP4, a gene which appears to be involved in specific stages of embryo development and is [36].

Proteasome component system proteins, such as proteasome lota, proteasome component C2, proteasome subunit RC-7 I, 26 S-proteasome and proteasome component C3 were down regulated by PGF2α. Proteasomal component systems are the main non-lysosomal proteolytic structures of the cells that participate in the elimination of abnormal proteins, short half-life proteins, and proteins controlling cell cycle [37]. During the process of cell differentiation, the level of proteasome expression and its localization varies. The proteasomal proteins can be intermediaries of ECM by contributing to the modulation of the cell cycle through the induction of proteasomal degradation of cyclin dependent kinase 2. Cell attachment to ECM components, such as fibronectin (FN), does not affect p21 mRNA levels, but the stability of the p21

### Table 4: Effect of PGF2α on the expression of genes associated with G-proteins, growth factors, chemokines and cytokines.

<table>
<thead>
<tr>
<th>Gene Bank number</th>
<th>Swiss protein number</th>
<th>Name</th>
<th>Function</th>
<th>Ratio ± 0.2</th>
<th>% decrease</th>
<th>Fold change</th>
</tr>
</thead>
<tbody>
<tr>
<td>X52498</td>
<td>P17246</td>
<td>Transforming Growth Factor β1</td>
<td>Growth factor</td>
<td>0.33</td>
<td>72</td>
<td>0.29 ± 0.15</td>
</tr>
<tr>
<td>M60921</td>
<td>P27049</td>
<td>Antiproliferative B-cell translocation gene 2 (BTG2)</td>
<td>Growth factor</td>
<td>0.28</td>
<td>51</td>
<td>0.25 ± 0.14</td>
</tr>
<tr>
<td>X61381</td>
<td>P26376</td>
<td>Interferon-induced protein</td>
<td>Chemokine</td>
<td>0.25</td>
<td>56</td>
<td>0.16 ± 0.14</td>
</tr>
<tr>
<td>L11586</td>
<td>Q64604</td>
<td>Leukocyte common antigen</td>
<td>Intracell. phosphatase</td>
<td>0.07</td>
<td>83</td>
<td>0.02 ± 0.02</td>
</tr>
<tr>
<td>Z22607</td>
<td>Q06826</td>
<td>Bone Morphogenetic protein 4 (BMP4)</td>
<td>Growth factor</td>
<td>0.34</td>
<td>78</td>
<td>0.02 ± 0.17</td>
</tr>
<tr>
<td>L12384</td>
<td>P26437</td>
<td>ADP Ribosylation factor 5</td>
<td>Intracellular transducer</td>
<td>0.18</td>
<td>67</td>
<td>0.06 ± 0.008</td>
</tr>
<tr>
<td>L12385</td>
<td>P26438</td>
<td>ADP Ribosylation factor 6</td>
<td>Intracellular transducer</td>
<td>0.18</td>
<td>83</td>
<td>0.07 ± 0.007</td>
</tr>
<tr>
<td>Z11558</td>
<td>Q63228</td>
<td>Gla maturation factor β (GMFB)</td>
<td>Growth factor</td>
<td>0.17</td>
<td>83</td>
<td>0.17 ± 0.07</td>
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<td>M30705</td>
<td>P14842</td>
<td>S-hydroxytryptamine receptor 2A (HTR2A)</td>
<td>G proteins</td>
<td>0.06</td>
<td>93</td>
<td>0.13 ± 0.03</td>
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<tr>
<td>S47609</td>
<td>P30543</td>
<td>Adenosine A2A receptor (ADORA2A)</td>
<td>G proteins</td>
<td>0.13</td>
<td>87</td>
<td>0.17 ± 0.08</td>
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<td>M91466</td>
<td>P29276</td>
<td>Adenosine A2B receptor (ADORA2B)</td>
<td>G proteins</td>
<td>0.17</td>
<td>83</td>
<td>0.22 ± 0.14</td>
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<tr>
<td>M20713</td>
<td>P08753</td>
<td>Guanine nucleotide-binding protein G α3 subunit</td>
<td>G proteins</td>
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<td>0.17 ± 0.07</td>
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<td>M17525</td>
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<td>G proteins</td>
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<td>M75153</td>
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<td>D85760</td>
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<td>Guanine nucleotide-binding protein α12 subunit</td>
<td>G proteins</td>
<td>0.15</td>
<td>85</td>
<td>0.15 ± 0.09</td>
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<tr>
<td>D12498</td>
<td>Q04589</td>
<td>Basic Fibroblast Growth Receptor factor 1</td>
<td>Growth factor</td>
<td>2.32</td>
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<td>2 ± 1.07</td>
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protein decreased [36]. Kinase activities such as ERKs, calcium/calmodulin kinase II and IV, SGK, and casein kinase can affect the ECM downstream or upstream [38]. On the other hand, ECM can regulate the availability of substrates, as well as factors or effectors downstream or upstream related with different cascades of signal transduction pathways. For example, proteins such as decorin are components of the ECM in many tissues and appear to be involved in matrix assembly [39]. Decorin can cause the rapid phosphorylation of EGF with the concurrent activation of mitogen-activated kinase protein kinase signal pathway. Via TGFβ, decorin can interact with the MAP kinases signal transduction pathways and cross talk with calcium/calmodulin-dependent kinase II [40], which in the cDNA array was down-regulated by PGF2α.

Crk-associated substrate (CAS) was down-regulated by PGF2α and is another gene related to the ECM. Active CAS can modulate changes in cell motility and gene expression by the various MAP kinase cascades, and modify the organization of the actin cytoskeleton [39]. Another gene down-regulated by PGF2α was SGK, which is a transcriptionally-regulated serine/threonine protein kinase with 45–55% homology to the catalytic domain of Akt/PKB protein kinase A [40]. Skg is expressed in decidual tissue and can be activated by the phosphoinositide 3-kinase pathway (PI3-Kinase) through PDK1-mediated phosphorylation [41]. Trafficking proteins such as 14-3-3 z/δ and ε are involved in the regulation of genes related to the ECM because it can block the activation cascade of signal transduction pathways [42].

Protein phosphatase type 2A (PP2A) was down regulated by PGF2α. The first and most important point of control of PP2A is at the transcriptional level. The increase of phosphatase activity corresponded with a decrease in the phosphorylation of cellular proteins in anchorage-dependent cells, but much lesser regulated in anchorage-independent cells [43].

It is well known that members of the proinflammatory cytokine family can induce MMP expression in numerous tissues [44]. Also, cytokines and chemokines, such as interferon inducible protein and leukocyte common antigen, are associated with the local induction of MMP expression in response to proinflammatory cytokines. The indirect action of these molecules through MMPs may aid to generate different changes in the endometrial stroma during maternal recognition of pregnancy [44].

We have also compared the pattern of gene expression in rat decidual tissue in vivo on day 12 of pseudopregnancy (when the decidua undergoes regression) with that of the PGF2α treatment in rat PDC in vitro (data not shown). Interestingly, we found a 49% coincidence in the genes that were down regulated in both experimental situations. This coincidence suggests a relationship between the physiological regression and reorganization of the decidual tissue that occurs on day 12 of pseudopregnancy, with the pattern of gene expression in rat PDC after PGF2α treatment. On the other hand, many of the genes whose expression was affected by PGF2α in the rat PDC, are expressed in the decidua during decidualization and implantation. This suggests a possible connection between the action of PGF2α and the physiology of the decidual tissue. Moreover, the ECM is an important component of the decidualization and implantation process. Any modification in its turnover or degradation could affect the formation of the decidua, blastocyst invasion, and the timing of decidual regression and reorganization. It is known that the expression of MMPs/TIMPs plays an important role in the control of implantation. If PGF2α silences or decreases the expression of genes related with the systems aforementioned, it also could affect tissue remodelling by directly modifying the proteases involved in this process, or indirectly by affecting the ECM turnover and degradation, important in the accumulation of a spongy mass of tissue around each embryo during decidualization [45]. Most probably, many of PGF2α’s effects on the expression of genes involved in ECM turnover are indirect. Alterations of genes involved in different signalling pathways such as ERK-1, CaMKII, PLCδ1 and G-protein subunits may impact the expression of a great number of transcripts.

In summary, our results show, for the first time, that pathophysiological concentrations of PGF2α have a severe impact on the expression of numerous genes associated with the turnover of the ECM in the rat decidua. Future investigation should corroborate the differentially regulated genes, at the level of the message, protein, and in some cases, such as for the metalloproteinases, at the level of activity of the proteins. These data will contribute to the design of future studies on a cluster of gene candidates as targets of PGF2α action in this endocrine tissue.

**Abbreviations**

TIMP2 = tissue inhibitor metalloproteinases 2; TIMP3 = tissue inhibitor metalloproteinases 3; PAI 1 = plasminogen activator inhibitor 1; uPA = urokinase type; tPA = tissue type plasminogen activator; TGFβ = transforming growth factor β; SGK = serum glucocorticoid kinase; ERK-1 = extracellular signal-regulated kinase-1; MAK = male germ cell-associated kinase; CAS = crk-associated kinase kinase; CamKII = calcium calmodulin kinase II; CamKIV = calcium calmodulin kinase IV; PLCδ1 = phospholipase C delta 1; BMP 4 = bone morphogenetic factor 4; ADPRF 5 = ADP ribosylation factor 5; bFGFRF1 = basic fibroblast growth factor receptor F1; HTR2A = 5-hydroxytryptamine receptor 2A; ADORA2A = adenosine receptor A2A;
ADORA2B = adenosine receptor A2B; GN-BPG α3 = guanine nucleotide-binding protein G α3; GN-BP α stimul. = guanine nucleotide-binding protein α stimulating; GN-BPG α1,3 Sub. = guanine nucleotide-binding protein α1,3 subunit

Authors’ contributions
EC and SFG carried out the decidualization, and primary decidual cell culture. EC isolated the RNA isolation, performed the CDNA array assay, the analysis of the results, and drafted the manuscript. GG conceived the study and edited the manuscript. All authors read and approved the final manuscript.

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References


