Research article

Mice expressing HLA-DQ6α8β transgenes develop polychondritis spontaneously

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Abstract

Relapsing polychondritis (RP) is a human autoimmune disease of unknown etiology in which cartilaginous sites are destroyed by cyclic inflammatory episodes beginning, most commonly, during the fourth or fifth decade of life. We have previously described collagen-induced polychondritis that closely mirrors RP occurring in young (6–8 weeks old) HLA-DQ6α8βαβ8βαβ transgenic Aβ0 mice, following immunization with heterologous type II collagen (CII).

We present evidence here that transgenic strains expressing the DQ6αβ8β transgene develop spontaneous polychondritis (SP) at the mouse equivalent of human middle age (4.5–6 months and 40–50 years old, respectively) and display polyarthritis, auricular chondritis and nasal chondritis – three of the most common sites affected in RP. Auricular chondritis in SP, like RP but unlike CII-induced polychondritis, exhibited a relapsing/remitting phenotype, requiring several inflammatory cycles before the cartilage is destroyed. Elevated serum levels of total IgG corresponded with the onset of disease in SP, as in RP and CII-induced polychondritis. No CII-specific immune response was detected in SP, however – more closely mirroring RP, in which as few as 30% of RP patients have been reported to have CII-specific IgG. CII-induced polychondritis displays a strong CII-specific immune response. SP also demonstrated a strong female preponderance, as some workers have reported in RP but has not observed in CII-induced polychondritis. These characteristics of SP allow for the examination of the immunopathogenesis of polychondritis in the absence of an overwhelming CII-specific immune response and the strong adjuvant-induced immunostimulatory influence in CII-induced polychondritis.

This spontaneous model of polychondritis provides a new and unique tool to investigate both the initiatory events as well as the immunopathogenic mechanisms occurring at cartilaginous sites during the cyclic inflammatory assaults of polychondritis.

Introduction

Relapsing polychondritis (RP) is a human autoimmune disease of unknown etiology. RP is relatively rare, affecting 3.5–4/1,000,000 people, with a slight female preponderance [1]. Typical onset occurs during mid-life years with a median age of diagnosis at 46.6 years, although cases have been reported in newborns and in individuals over the age of 80 [2]. The most prominent manifestations of RP are auricular chondritis, polyarthritis and saddle nose deformation, with auricular chondritis being the most common presenting symptom [1,2]. Any other cartilaginous tissue may also be affected in RP, including the eyes, the trachea and the respiratory tract [2,3]. Other tissues can also be affected since some RP patients have demonstrated renal involvement and skin lesions [2,4]. Autoantibodies specific for type II, type IX and type XI collagen and for matrilin-1 [5-8] have all been detected in sera from RP patients, supporting RP as an autoimmune disease.

We have previously described a murine model of induced polychondritis that mimics many characteristics of RP [9,10].
Young mice (6–8 weeks of age) expressing the HLA-DQ6αβtg molecule, either the DQ6αβtg transgenic (tg) or DQ6αβtg β transgenic strains, develop polychondritis following immunization with an emulsion of type II collagen (CII) and Complete Freund's Adjuvant. Induced polychondritis is characterized by a single acute inflammatory event, by auricular chondritis or by polyarthritis, followed by the destruction of the cartilage in both the peripheral joints and ears, and by pannus formation. Neither of the parental strains, DQαβtg mice or DQ8αβtg mice, develop polyarthritis, although DQ8αβtg mice do develop collagen-induced arthritis [9,10]. Hansson and colleagues [11] have also recently described a model of polychondritis, induced in DBA mice and in Lew congenic rats by immunization with bovine matrilin-1 in Complete Freund's Adjuvant. In that model, there is inflammation of the trachea and respiratory tract, followed by their destruction; however, the ears, joints and other cartilaginous sites are not affected.

We present evidence that when mice expressing the DQ6αβtg transgene and lacking both endogenous H2-A and H-2E expression, either DQ6αβtg or DQ8αβtg mice, reach middle age (between 4 and 6 months of age), they develop spontaneous polychondritis (SP), with inflammatory events occurring at the ears, the peripheral joints and the nose. Auricular chondritis in SP displayed a relapsing/relenting phenotype, more closely mirroring human RP than induced polychondritis. This model occurs in the absence of overt stimulation by an exogenous antigen such as CII, or adjuvant, or by artificially induced trauma to any of the sites of inflammation, and provides an excellent model that mirrors the human disease more closely than any previously described model of RP.

Materials and methods

Mice
The generation of mice expressing both HLA-DQ6αβ (DQA1*0103/DQB1*0601) and HLA-DQ8αβ (DQA1*0301/ DQB1*0302) transgenes in the absence of endogenous MHC class II expression (Aβ0) was previously described [9], and the mice were initially provided by Dr Chella David (Mayo Clinic, Rochester, MN, USA). Briefly, the transgenes were introduced into the H2f background crossed with Aβ0 mice, resulting in tg mice expressing the DQ transgene and lacking both H2-A and H2-E expression. All breeding occurred in laminar flow hoods to maintain a relatively pathogen-free environment. Retired breeders and mice aged greater than 6 months were moved to clean conventional rooms in the Center for Biomedical Research. Mice were given rodent chow and water ad libitum. All studies were approved by the University of North Dakota Institutional Animal Care and Usage Committee. Mice were bled retro-orbitally; the blood was spun for 10 minutes at 12,000 × g and the serum was decanted for analysis of IgG specificity.

Total IgG levels
Total serum IgG levels were determined by ELISA as previously described [12]. Briefly, serum from experimental mice was diluted fourfold, 1/1,000–1/64,000, in coating buffer (1 M Na2CO3 + 1 M NaHCO3 + 1 M NaN3), and 200 μl was added to Nunc Maxisorb microtiter plates in duplicate and incubated at room temperature for 2 hours. Plates were washed three times with PBS/Tween as in CII antibody, and then 100 μl/well goat anti-mouse immunoglobulin (Sigma-Aldrich, St Louis, MO, USA) was added and plates were incubated for 2 hours at 37°C. Plates were washed three times with PBS/Tween, and then 200 μl/well pNpp substrate (Sigma-Aldrich) was added and incubated for 1 hour at 37°C to visualize the reaction. The colorimetric change of each well was determined at an optical density of 405 nm on a Thermomax Microplate Reader (Molecular Devices, Sunnyvale, CA, USA), and the data were analyzed using the SOFTmax analytical software program (Molecular Devices) and Microsoft Excel.

Type II collagen-specific antibody levels
One hundred microliters of collagen solution (Arthrogen-CIA mouse type II collagen, 5 mg/ml) diluted in 1 × collagen dilution buffer as per the manufacturer’s instructions (Chondrex, Redmond, WA, USA) was added to Nunc Maxisorb microtiter plates (NNI, Rochester, NY, USA) and incubated overnight at 4°C. Plates were washed three times with PBS/0.5% Tween 20, were blocked by adding 1% BSA in PBS (blocking buffer) for 2 hours at room temperature and were then washed again three times with PBS/Tween. Experimental serum was diluted fourfold, 1/1,000–1/64,000, in blocking buffer/Tween, and then 200 μl was added to the plates in duplicate.

Titration of a high-titer CII-specific IgG-positive control and a CII-negative serum control were run with each plate. Sera were incubated for 2 hours at 37°C and were washed three times with PBS/Tween. CII-specific mouse IgG was detected with peroxidase-conjugated goat anti-mouse IgG (Sigma-Aldrich) for 1 hour at 37°C. The plates were again washed three times with PBS/Tween and the reaction visualized with pNpp substrate (Sigma-Aldrich). The colorimetric change of each well was determined at an optical density of 405 nm on a Thermomax Microplate reader, and the data were analyzed using the SOFTmax analytical software program (Molecular Devices) and Microsoft Excel.

Detection of matrilin-1 antibodies
The presence of matrilin-1-specific IgG was determined in sera from representative SP mice, in sera from young and middle-aged naïve DQ6αβtg β mice and in sera from CIA-immunized young DQ6αβtg β mice by western blot as previously described [8]. Briefly, matrilin proteins were extracted from bovine epiphyseal cartilages, were resolved by SDS-PAGE and were transblotted to polyvinylidene difluoride membrane (BioRad, Richardon, CA, USA) for western blot analysis. The
western blot analysis was performed with mouse serum diluted 1:100 and probed with an alkaline phosphatase-conjugated goat anti-mouse IgG Fc (Jackson ImmunoResearch, Avondale, PA, USA). The blots were developed using 5-Bromo-4-chloro-3-indolyl phosphate/nitroblue tetrazolium.

**Histological analysis**

Outer ears displaying clinical chondritis or outer ears from undiseased normal controls were surgically removed from mice anesthetized with Isoflurane (Phoenix Pharmaceutical Inc., St. Joseph, MO, USA) at various time points in the course of disease. Ears were embedded in paraffin, were sectioned, were mounted onto slides and were stained with hematoxylin and eosin. Front and hind limbs were removed and dissected from euthanized mice at the conclusion of the experiment. Joints were decalcified and embedded in paraffin. Sections were cut, mounted and stained with hematoxylin and eosin. Sections of both ears and joints were also stained with Safranin O or Toluidine Blue to determine the presence of mast cells. All sections were assessed in a blinded fashion by Dr. Maryann Sens (Department of Pathology, UNDSMHS, Grand Forks, ND, USA) for histopathological changes and for cellular infiltration.

**Statistical analysis**

Statistical analysis was carried out using the Mann–Whitney rank sum test for nonparametric tests. P < 0.05 was considered statistically significant unless otherwise noted.

**Results**

**Spontaneous onset of polyarthritis**

The incidence of SP in both DQ6αβ8αβ tg and DQ6αββ tg strains of mice is summarized in Table 1. DQ6αβ8αβ tg mice developed spontaneous polyarthritis at an incidence of about 80%, with 31% developing auricular chondritis. DQ6αββ tg mice developed spontaneous polyarthritis at an incidence of 76%, with 32% developing auricular chondritis (Figure 1). There was a strong female predisposition to developing SP in both DQ6αββ tg expressing strains: 3.8:1 and 4.4:1 female: male ratios in the DQ6αβ8αβ tg and DQ6αββ tg strains, respectively. Neither of the parental strains, DQ6αββ tg mice or DQ8αββ tg mice, developed SP. As we have reported elsewhere [13], however, DQ8αββ tg mice developed spontaneous polyarthritis at an incidence of 80% with a few mice (<1%) displaying inflammatory events at other sites.

The onset of spontaneous polyarthritis was observed from 4 to 11 months of age with a mean ± standard error of the mean onset of 5.8 ± 0.1 months in the DQ6αβ8αβ tg strain and of 5.6 ± 0.2 months in the DQ6αββ tg strain. Female mice developed disease at a significantly earlier time points in both strains, compared with males (Figure 1). Spontaneous auricular chondritis was delayed slightly more than 1 month from the onset of polyarthritis in both strains (7.2 ± 0.4 and 7.4 ± 0.2 for DQ6αβ8αβ tg and DQ6αββ tg strains, respectively), and independent of gender. The disease was clinically not as severe as with the induced polychondritis model. These mice, however, displayed a relapsing/remitting pattern of inflammation of their ears in which the ears would become swollen and red (Figure 2) for a period of 1–2 weeks, after which the inflammation would abate for a period of 1–2 weeks. This pattern of cyclic inflammation more closely resembles the manifestation of auricular chondritis found in human RP patients. After a period of alternating relapsing and remitting phases, the ear eventually becomes a chronic ‘cauliflower’ ear and is reduced to a small fibrous mass as shown in Figure 2. Histological analyses of ears at various time points during these inflammatory cycles demonstrated a corresponding pattern of infiltration and a retreat of mononuclear cells during acute and remitting phases (data not shown).

The digits, the paws and the distal joints were affected in both DQ6αβ8αβ tg mice and DQ6αββ tg mice displaying an arthritic state with redness and swelling; compared with young or middle-aged nonarthritic DQ6αββ tg expressing mice (Figure 3). The severity of polyarthritis was equivalent in both strains with SP, and was comparable with the polyarthritis observed in induced polychondritis (data not shown). Histological examination of sections of arthritic joints of the middle-aged spontaneous mice demonstrated a correlation between cellular infiltration, predominantly mononuclear cells including mast cells (Figure 4), and the severity of polyarthritis. Sections from joints from young, nondiseased mice and from middle-aged, nonarthritic mice demonstrated normal joint architecture with no signs of inflammation or infiltration of cells. Interestingly, the level of inflammation and the numbers of cellular infiltrates appeared to be lower in spontaneous mice than that observed in collagen-immunized mice (Figure 4).

A small percentage (<25%) of the DQ6αββ tg expressing mice with SP also displayed inflamed noses (Figure 5). Histological analysis has shown that there is a massive infiltration of mononuclear cells (data not shown). Eyes of mice with SP were compared with age-matched unaffected animals and were examined for ocular involvement, including conjunctivitis, keratitis, episcleritis or uveitis associated with RP. No changes were observed that correlated with disease, although some changes related to general aging were noted (data not shown). The kidneys were also examined histologically and no cellular infiltrate was observed (data not shown).

Previous studies in human rheumatoid arthritis and RP patients and the collagen-induced arthritis models have shown antibody responses to a variety of collagens, particularly CII [5,7], as well as cartilage matrix proteins such as matrilin-1 [8], especially during the acute phase of the disease. As CII-induced murine models of both RP and rheumatoid arthritis are dependent on both exogenous CII and adjuvant, it was of particular interest whether antibody levels were elevated in SP
and what antibody specificities were present in the lack of the strong but artificial CII-specific response.

ELISAs were performed to determine total and anti-CII-specific IgG antibody levels during the disease course. All mice displaying SP also displayed a significant increase in total IgG levels, similar to the total IgG response observed in induced polychondritis in young CII-immunized mice (Figure 6). None of the spontaneous mice, however, displayed any detectable CII-specific antibodies (Figure 6). To determine whether mice developing SP displayed an antimatrilin-1 antibody response, as detected in RP patients [8], matrilin proteins were separated by SDS-PAGE, were transferred to a polyvinylidene difluoride membrane and were blotted with DQ6αβ8αβ experimental sera.

Figure 7 shows a representative blot in which sera from mice were blotted against either matrilin proteins or CII. A strong IgG response to matrilin-1 was detected in sera from mice developing SP, but these mice lacked any CII-specific IgG.

Conversely, young, CII-immunized DQ6αβ8αβ tg sera displayed antibodies to CII but lacked significant antibodies to matrilin-1. Sera from middle-aged DQ6αβ8αβ tg mice not exhibiting any clinical signs of polychondritis did have low levels of matrilin-1-specific IgG, yet significantly lower than those with SP. These data suggest that CII is probably not involved in the autoimmune reaction responsible for SP, and suggest that matrilin-1 may be at least one of the putative autoantigens involved in either the induction or progression of SP disease.

**Discussion**

Few spontaneous models of arthritic disease have been observed [14-16], and none have as yet described all the manifestations necessary to be considered a spontaneous model of relapsing polychondritis – which is characterized not only by polyarthritis, but also by involvement of the ears, the nose and possibly the trachea. Hansson and colleagues have described a chondritic model in which matrilin-1 immunization induces involvement of the respiratory tract but in which peripheral joints and ears are unaffected [11].

![Figure 1](image-url)  
**Figure 1**

Incidence and onset of spontaneous polychondritis in female versus male DQ6αβ8β transgenic expressing mice. The onset of auricular chondritis and polyarthritis was determined by visual inspection and was confirmed by histological analyses as described in Table 1. Females (black shading) were more highly susceptible to both auricular chondritis and polyarthritis, and displayed clinical disease earlier than males (grey shading).
We describe herein a model of spontaneous autoimmune polychondritis that occurs in mice expressing the DQ6α8β transgene without the exogenous stimulation by antigen or adjuvant and the possible interference those substances may add. The mean onset of disease occurs about 6 months of age. Although difficult to directly correlate mouse years with human years, this places the onset of disease approximately at middle age, or near the fourth or fifth decade of life in human years, matching the mean onset of RP in humans. There is a strong female preponderance, which correlates with some reports of human relapsing polychondritis [1] and autoimmune diseases in general. This would suggest that the gender-related factors that make females more prone to autoimmune responses may also be occurring in this newly identified model, and thus may be better understood. It is of note that enhanced female susceptibility is not observed in the CII-induced model of polychondritis or arthritis.

Human studies have shown that as many as two-thirds of RP patients do not develop antibodies to CII during the course of disease [17], correlating to the finding that there was not a CII-specific immune response during SP. There are, however, significant total IgG responses in both SP and RP, suggesting a
role for antibodies in the pathogenesis of disease. In addition, there is an increased infiltration of mast cells into the synovium and ears during SP, which also correlates with other recent reports suggesting a pathogenic role for the elevated IgG through mast cells in RA and animal models of arthritis [18-21]. We are currently investigating the contribution the mast cells infiltrating into the cartilaginous tissues have toward the pathogenesis of SP. In addition, a strong antibody response to matrilin-1 is observed during SP, raising the possibility that matrilin-1 may be the putative autoantigen in the initiation of SP. This would correspond with an antimatrilin-1 antibody response observed in human RP patients [8] and with the ability of heterologous matrilin-1 to induce a polychondritis-like disease in mice [11].

There has been significant interest in another putative autoantigen, the ubiquitous protein glucose-6-phosphate-isomerase (G6PI), because antibodies specific for G6PI have been found...
Correlations have been made between the HLA genotype and the genetic predisposition to many autoimmune diseases, including rheumatoid arthritis (HLA-DR4, DOB), insulin-dependent diabetes mellitus (DR4, DOB), thyroiditis (DR3) and multiple sclerosis (DR4) [25]. The occurrence of SP in a mouse expressing the DQ6ββ tg expressing strains displayed an overall increase in the levels of TGF-β, regardless of the presence of disease, as compared with controls (naïve B10.T(6R) mice), with many samples reading high and out of range (Bradley and Cafruny, personal communication). It is unclear how this finding correlates with the immunoregulatory role of TGF-β in arthritis. It is possible, however, that the elevated TGF-β levels represent an active attempt by these mouse strains to downregulate the vigorous (auto)immune response. Alternatively, the TGF-β serum levels may represent another form of dysregulation, such as the role of TGF-β-specific antibodies in lupus-like diseases [24].

Treatment with transforming growth factor beta (TGF-β) has been shown recently to provide some protective effect against collagen-induced arthritis in young, susceptible mice, although the effects are not long-lasting. In aging mice, however, treatment with TGF-β has been shown to block induction of arthritis altogether [23]. DQ6ββ tg expressing mice displayed an overall increase in the levels of TGF-β, regardless of the presence of disease, as compared with controls (naïve B10.T(6R) mice), with many samples reading high and out of range (Bradley and Cafruny, personal communication). It is unclear how this finding correlates with the immunoregulatory role of TGF-β in arthritis. It is possible, however, that the elevated TGF-β levels represent an active attempt by these mouse strains to downregulate the vigorous (auto)immune response. Alternatively, the TGF-β serum levels may represent another form of dysregulation, such as the role of TGF-β-specific antibodies in lupus-like diseases [24].

**References**